Mycobacterium tuberculosis Bacteremia among Acutely Febrile Children in Western Kenya

Patricia B. Pavlinac,* Jaqueline M. Naulikha, Grace C. John-Stewart, Frankline M. Onchiri, Albert O. Okumu, Ruth R. Sitati, Lisa M. Cranmer, Erica M. Lokken, Benson O. Singa, and Judd L. Walson

Department of Global Health, University of Washington, Seattle, Washington; Department of Pediatrics, University of Washington, Seattle, Washington; Department of Epidemiology, University of Washington, Seattle, Washington; Department of Medicine, University of Washington, Seattle, Washington; Kenya Medical Research Institute, Centre for Clinical Research, Nairobi, Kenya; Kenya Medical Research Institute (KEMRI)/CGHR Centre for Global Health Research, Kisumu, Kenya; Department of Pediatria, Emery University School of Medicine and Children's Healthcene of Adjusta, Alunta, Coopring

Department of Pediatrics, Emory University School of Medicine and Children's Healthcare of Atlanta, Atlanta, Georgia

Abstract. In children, Mycobacterium tuberculosis (M. tuberculosis) frequently disseminates systemically, presenting with nonspecific signs including fever. We determined prevalence of M. tuberculosis bacteremia among febrile children presenting to hospitals in Nyanza, Kenya (a region with high human immunodeficiency virus (HIV) and M. tuberculosis prevalence). Between March 2013 and February 2014, we enrolled children aged 6 months to 5 years presenting with fever (axillary temperature $\geq 37.5^{\circ}$ C) and no recent antibiotic use. Blood samples were collected for bacterial and mycobacterial culture using standard methods. Among 148 children enrolled, median age was 3.1 years (interquartile range: 1.8–4.1 years); 10.3% of children were living with a household member diagnosed with M. tuberculosis in the last year. Seventeen percent of children were stunted (height-for-age z-score < -2), 18.6% wasted (weight-for-height z-score < -2), 2.7% were HIV-infected, and 14.2% were HIV-exposed uninfected. Seventeen children (11.5%) had one or more signs of tuberculosis (TB). All children had a Bacille Calmette-Guerin vaccination scar. Among 134 viable blood cultures, none (95% confidence interval: 0–2.7%) had Mycobacterium isolated. Despite exposure to household TB contacts, HIV exposure, and malnutrition, M. tuberculosis bacteremia was not detected in this pediatric febrile cohort, a finding consistent with other pediatric studies.

INTRODUCTION

Tuberculosis (TB) is a common cause of morbidity and mortality among children living in sub-Saharan Africa (SSA).^{1,2} While *Mycobacterium tuberculosis* (*M. tuberculosis*) typically manifests as isolated pulmonary disease in immune-competent adults, in young children and in those who are immunosuppressed (with HIV or severe malnutrition), *M. tuberculosis* is more likely to disseminate.^{3,4} Disseminated *M. tuberculosis* often presents with nonspecific signs and symptoms, including fever, weight loss, and general malaise, similar to the signs and symptoms of many other infections. Disseminated *M. tuberculosis* in children is associated with a very high case fatality rate, exceeding 50% in some reports.^{5–7} Although prompt therapy likely reduces mortality, mycobacterial culture is not available and is cost-prohibitive in many settings leading to delays in diagnosis.

Among febrile, hospitalized adult populations in SSA, *M. tuberculosis* is a leading cause of bloodstream infection, with prevalence estimates ranging from 5.5% to 23.4%.^{5,8–12} In this population, human immunodeficiency virus (HIV)-associated immunosuppression has emerged as the most important risk factor for *M. tuberculosis* bacteremia. However, studies assessing *M. tuberculosis* bacteremia in children are limited and the prevalence of *M. tuberculosis* bacteremia in children with acute febrile illness has not been described.^{13–15} Given the difficulty in establishing the diagnosis of *M. tuberculosis* bacteremia in many settings and the need for prompt initiation of anti-tuberculosis therapy to reduce mortality, we sought to determine the frequency and correlates of *M. tuberculosis* bacteremia in Kenyan children aged 6 months to 5 years presenting with fever to one of three western Kenya hospitals.

METHODS

Consecutive children aged 6 months to 5 years, without recent antibiotic use (defined as documented antibiotic use within the last 24 hours other than prophylactic cotrimoxazole for HIV exposure) were enrolled in a cross-sectional study evaluating bacterial causes of bloodstream infections. Children were enrolled from three health facilities (Kisii Teaching and Referral Hospital, Homa Bay District Hospital, and Migori District Hospital) in the Nyanza region of Kenya where the estimated adult prevalence of HIV and TB are 15.1% and 0.6%, respectively.^{16,17} Children enrolled between February and August 2013 and between January and March 2014 were asked to provide additional blood for mycobacterial blood culture. The gap in enrollment in this substudy was due to unavailability of the appropriate mycobacteria culture bottles for the time period between September 2013 and December 2013. Written informed consent was obtained from primary caregivers of enrolled children. Study clinical officers examined children according to the World Health Organization (WHO) Integrated Management of Childhood Illness (IMCI) guidelines, obtained height and weight, and assessed for presence of a Bacille Calmette-Guerin (BCG) scar.¹⁸ Height-for-age (HAZ), weight-for-height (WHZ), and weight-for-age z-scores (WAZ) were calculated using the 2006 WHO reference populations for children under 5 years.¹⁹ Stunting and wasting were defined as HAZ < -2 and WHZ < -2, respectively. A standardized questionnaire was used to obtain clinical history, signs and symptoms from the physical examination, and sociodemographic information.

Up to 5 mL of blood was collected aseptically and separated for bacterial culture, HIV testing, malaria testing, and up to 3 mL for mycobacterial culture. Maximum allowable blood draw volumes (2 mL/kg) were set based on those used at Seattle Children's Hospital and additional blood for mycobacterial blood culture was drawn only when an additional 1–3 mL could be drawn from the child. Children were tested

^{*}Address correspondence to Patricia Bearden Pavlinac, Department of Global Health, University of Washington, 325 9th Avenue, Box 359931, Seattle, WA 98104. E-mail: ppav@u.washington.edu

for HIV using antibody testing (Abbott Determine[™] rapid test kit and confirmed using Uni-Gold[™]) or HIV RNA polymerase chain reaction if < 18 months of age. Maternal HIVstatus was ascertained by self-report if the mother reported being HIV-infected and confirmed by antibody testing if the mother was unsure of her infection status or reported being HIV uninfected. Malaria was assessed at the study site using both rapid diagnostic testing (RDT) (Paracheck Pf® Orchid Biomedical Services, India) and microscopy.

For mycobacteria blood culture, 1-3 mL of blood inoculated in Bactec[™] Myco F/lytic bottles were shipped daily to the nearby Kenya Medical Research Institute (KEMRI)/Center for global health research (CGHR) TB laboratory in Kisian, Kenya where they were immediately placed in a Bactec 9120 machine for incubation. The transit time to the laboratory was less than 24 hours. Flagged bottles were removed, mixed gently, and subcultured to blood agar plates and Ziehl-Neelsen (ZN) smears. Positive ZN smears were subcultured onto Löwenstein-Jensen solid medium and evaluated for an additional 3 weeks. Flagged bottles that were ZN negative and positive on brain heart infusion agar were deemed contaminated. Unflagged bottles at 42 days were considered negative. Bacterial blood cultures were performed at the U.S. Army Medical Research Unit Microbiology Hub laboratory in Kericho, Kenya. Methods and results of bacterial culture for the parent study are presented elsewhere.²⁰

Frequencies and percentages of clinical and sociodemographic characteristics were calculated and the 95% confidence interval (CI) around the M. tuberculosis bacteremia prevalence estimated assuming a binomial distribution. Analyses were conducted using Stata 11.1 (Stata Corp., College Station, TX). The University of Washington Institutional Review Board and the KEMRI Ethical Review Committee approved the study.

RESULTS

Of 375 children enrolled, 148 (39.5%) agreed to participate in the M. tuberculosis substudy and met weight standards for maximum blood draw quantity after blood for bacterial culture, malaria, and HIV testing was collected. Enrolled children were a median of 3.1 years of age (interquartile range [IQR]: 1.8-4.1 years), 54.7% were male, and 43.9% lived in a household with two or more people per room (Table 1). All children had a BCG scar. Reflective of the underlying population of children seeking care for acute illness in the study settings, malnutrition was relatively common; 17.0% of children were stunted and 18.6% wasted. In addition, 2.7% of children were HIV-infected and 14.2% were HIV-exposed uninfected. The median temperature at presentation was 38.6°C (IQR: 38.2-39.3°C). One third of the children (28.4%) presented with at least one IMCI-defined general danger sign and 11.5% had signs and symptoms suggestive of M. tuberculosis (5.4% with 2 or more weeks of night sweats, 5.4% with 2 or more weeks of cough, and 1.4% with 1 or more weeks of fever). Over a third (36.5%) of children had a positive malaria RDT result and 36.4% had parasitemia.

Fourteen children (10.3%) lived in the same household as someone diagnosed and treated for TB in the last year. The identified household TB case was the mother for four children, the father for three children, both the mother and father for one child, and other household members for the remaining

TABLE 1 Characteristics of enrolled children (N = 148)

Characteristic	п	
	Median (IQR)	
Sociodemographic		
Median age (years)	3.1 (1.8-4.1)	
Male	81 (54.7%)	
Site		
Kisii	72 (48.7%)	
Homa Bay	67 (45.3%)	
Migori	9 (6.1%)	
Monthly household income	72 (48.7%)	
< 5,000 Kenyan Shilling		
Crowding*	65 (43.9%)	
Parent/caregiver married	126 (85.2%)	
Clinical history	· · · · ·	
Household TB [†]	14 (10.3%)	
BCG scar	148 (100%)	
Median number of months	6 (4-6)	
exclusively breast-fed		
Currently breast-feeding [‡]	24 (55.8%)	
Hospitalized in the last year	15 (10.1%)	
Malaria§	54 (36.5%)	
HIV-exposed uninfected	20 (14.2%)	
HIV-infected¶	4 (2.7%)	
HIV-associated immunosuppression**	1 (25%)	
Enrolled in care	1 (25%)	
Taking cotrimoxazole	1 (100%)	
Taking antiretroviral therapy	0 (-)	
Clinical presentation		
Any IMCI danger sign	42 (28.4%)	
Unable to drink or breast-feed	16 (10.8%)	
Excessive vomiting	30 (20.3%)	
Convulsions	6 (4.1%)	
Lethargy/unconscious	3 (2.0%)	
Any pneumonia sign	25 (17.2%)	
Chest in-drawing	0 (-)	
Stridor in calm child	0 (-)	
Fast breathing	25 (17.2%)	
Any TB sign	17 (11.5%)	
2 + weeks of night sweats	8 (5.4%)	
2 + weeks of cough	12 (8.1%)	
1 + week of fever	2 (1.4%)	
Stunted ^{††} (HAZ < -2)	24 (17.0%)	
Wasted \ddagger (WHZ < -2)	26 (18.6%)	
Acute malnutrition (MUAC < 12.5 cm)	7 (4.7%)	

BCG = Bacille Calmette-Guerin; HAZ = height-for-age z-score; HIV = human immuno-deficiency virus; ICMI = Integrated Management of Childhood Illness; IQR = interquartile range; RDT = rapid diagnostic test; TB = tuberculosis; WHZ = weight-for-height z-score. * ≥ 2 people per room living in house.

⁺ Member of household diagnosed or treated for TB in last year. [±] Among 43 children under ≤ 24 months of age.

 Among 43 children under ≤ 24 months of age.
§ Diagnosed by malaria RDT or smear microscopy (54/148 positive by RDT; 52/143 positive by microscopy

||Among 141 children who were HIV-uninfected and accompanied by their biological mother.

Three out of four children were diagnosed with HIV as part of this study.

Three out of our chindra were dangeded with thr as particle has study. ** Defined in terms of CD4% (age ≤ 11 months: < 25%, 12–35 months: < 20%, 36+ months: < 15%) or, in absence of CD4% data, in terms of CD4 count (age ≤ 11 months: < 1,500 cells/mm³, 12–35 months: < 750 cells/mm³, 36⁺ months < 350 cells/mm³). † †HAZ less than -6 and greater than 6 were considered to be implausible values and

set to missing

^{‡‡}WHZ less than −6 and greater than 6 were considered to be implausible values and set to missing

six. Only one of the 14 (7.1%) TB-exposed children was being treated with isoniazid preventive therapy per caregiver report.

A bacterial pathogen was isolated from blood culture in 5 (3.4%) of the 148 children, 4 had non-typhoidal Salmonella and 1 Staphylococcus auerus, and 3 (2.0%) had a potential bacterial contaminant (Staphylococcus epidermidis was isolated in one sample and Micrococcus in two). Fourteen of the mycobacterial cultures (9.5%) were not evaluable for mycobacteria due to overgrowth of bacteria or contamination, 5 (35.7%) of which were from children who had positive bacterial cultures (2 Salmonella O Poly A, 1 Salmonella choleraesuis, and 2 Micrococcus spp.). No Mycobacterium (0%) was identified in the remaining 134 samples (95% CI: 0-2.7%).

DISCUSSION

We did not identify *M. tuberculosis* from the bloodstream of febrile children presenting to several hospitals in western Kenya despite the relatively high prevalence of HIV, M. tuberculosis, and malnutrition in this region.^{17,21} Several studies in Africa have reported that in acutely ill HIVinfected adults, M. tuberculosis bacteremia is common and is found primarily in those with profound immunosuppression and severe illness.^{9,10} The lack of *M. tuberculosis* bacteremia identified in children observed in this study is consistent with previous pediatric studies, which have demonstrated the absence or extremely low prevalence of M. tuberculosis bacteremia in children.¹³⁻¹⁵ Although our study included relatively few children with HIV, a known risk factor for disseminated TB, a recent study in Tanzania also reported no evidence of M. tuberculosis bacteremia among 93 HIVinfected infants and children with frequent severe immunosuppression (over 60%) and presenting with severe illness; however, only 25% had fever.13

Although young age is a risk factor for the most severe forms of TB, including disseminated TB, available evidence suggests that M. tuberculosis bacteremia is uncommon among young children.^{4,22} For bacterial bloodstream infections, blood volume and number of samples correlate with yield.^{23,24} Therefore, it is possible that *M. tuberculosis* bacteremia is missed in children due to inadequate detection tools and blood volume limits. However, it should be noted that M. tuberculosis bacteremia was identified in a number of adult studies that reported inoculation volumes of 5 mL or less.^{10,25,26} Children with TB are also more likely to have paucibacillary disease, which, when combined with low blood volumes, may also limit the ability to detect circulating TB.²⁷ Given these diagnostic limitations, more sensitive methods may be needed to detect *M. tuberculosis* bacteremia in children, although molecular methods have had mixed diagnostic utility in adults.²⁸⁻³⁰ Widespread use of BCG vaccine may be another reason why TB bacteremia is rarely seen in children. BCG vaccine has demonstrated protection against TB meningitis and disseminated TB during infancy, but this protection wanes by adulthood.^{31–34}

This study had several notable strengths. The inclusion of young, febrile children from areas of high M. tuberculosis and HIV prevalence yielded a cohort with plausible risk for M. tuberculosis bacteremia. The use of standardized mycobacterial culture techniques in the CGHR laboratory was also a strength. The study also had several limitations. First, we included all children with fever, including those without HIV, malnutrition, or immunosuppression, who may have had lower risk for *M. tuberculosis* bacteremia. However, because the clinical presentation of children with M. tuberculosis bacteremia is largely unknown, this nonspecific inclusion criteria could have led to identification of novel signs and symptoms of *M. tuberculosis* bacteremia in children. By excluding children with recent antibiotic use (because of the parent study's goal of identifying bacterial causes of bloodstream infections) we may have excluded some children who were failing antibiotic therapy due to *M. tuberculosis* disease. *M. tuberculosis* bacteremia may be more likely to be detected in a cohort restricted to HIV-infected, immunosuppressed, and febrile children who fail antibiotic therapy.³⁵ Although 3 mL was the targeted amount of blood to be inoculated, as little as 1 mL was accepted in cases when the volume blood limits prohibited collecting the optimal volume. Actual blood volume collected was not recorded, however, the importance of obtaining 3 mL when possible was emphasized throughout all study trainings. A final limitation is that potentially useful clinical data (such as the presence of lymphadenopathy or hepatosplenomegaly) from the physical examination were not captured, prohibiting reporting these potential indicators of TB in children.

In this study of febrile children in areas of high HIV and TB prevalence, we did not find evidence of *M. tuberculosis* bacteremia. Although children make up as much as 21% of the global tuberculosis burden, detection of *M. tuberculosis* disease remains a major challenge in this population.^{36,37} This study adds to the body of evidence suggesting *M. tuberculosis* bacteremia is either very rare or undetectable in pediatric populations. Given the increased risk and high case fatality of disseminated TB in children, more sensitive methods for detection are needed to exclude *M. tuberculosis* bacteremia in children presenting with fever in high TB- and HIV-prevalence settings.

Received May 15, 2015. Accepted for publication July 13, 2015.

Published online August 31, 2015.

Acknowledgments: We would like to thank all of the participants and clinics that played a role in this study. We would also like to acknowledge Doreen Rwigi, Alfred Odiwuor, and other staff of the University of Washington/Kenya Medical Research Institute collaboration, without whom this work would not be possible. This research and publication were made possible with support from the University of Washington Center for AIDS Research (CFAR), an NIH funded program (P30 AI027757), which is supported by the following NIH Institutes and Centers (NIAID, NCI, NIMH, NIDA, NICHD, NHLBI, NIA). Additionally we would like to thank the CFAR-supported TB/HIV group for Collaboration of Research Efforts (TB/HIV CORE), the Healthy Growth and Development Core (UW Global Center for Integrated Health of Women, Adolescents and Children [Global WACh]) and the Kenva Research Program for their support during the preparation of this proposal and manuscript. We would also like to thank Solomon Mpoke, the director of the Kenya Medical Research Institute, for his support. The findings and conclusions in this paper are those of the authors and are not to be construed as official, or as reflecting true views of the Kenya Medical Research Institute, the University of Washington, or other affiliated institution.

Financial support: This study was made possible by funding from the National Institute of Health [grant number U19-A2090882] and the Firland Foundation. The funders of the study had no role in the study design, data collection, data analysis, data interpretation, or writing of the manuscript. The study was designed and implemented by the study investigators and the investigators conducted the analysis and prepared the manuscript. Patricia B. Pavlinac was supported by the University of Washington STD/AIDS Research Training Program [grant number T32-AI007140]. Grace C. John-Stewart is supported by a National Institute of Health mentoring award (grant number K24-HD054314). Lisa M. Cranmer is supported through the Pediatric Scientist Development Program (NICHD K12 HD000850, American Academy of Pediatrics, and American Pediatric Society).

Authors' addresses: Patricia B. Pavlinac, Department of Global Health, University of Washington, Seattle, WA, E-mail: ppav@uw .edu. Jaqueline M. Naulikha, Department of Pediatrics, University of Washington, Seattle, WA, and Kenya Medical Research Institute, Centre for Clinical Research, Nairobi, Kenya, E-mail: jngarayi@gmail.com. Grace C. John-Stewart, Departments of Medicine, Pediatrics, Epidemiology, and Global Health, University of Washington, Seattle, WA, E-mail: gjohn@uw.edu. Frankline M. Onchiri and Erica M. Lokken, Department of Epidemiology University of Washington, Seattle, WA, E-mails: fonchiri@uw.edu and erica.lokken@gmail.com. Albert O. Okumu and Ruth R. Sitati, Kenya Medical Research Institute/ Centre for Global Health Research, Kisumu, Kenya, E-mail: AOchiengOkumu@kemricdc.org and rsitati@kemricdc.org. Benson O. Singa, Kenya Medical Research Institute, Centre for Clinical Research, Nairobi, Kenya, E-mail: singabo2008@gmail.com. Lisa M. Cranmer, Department of Pediatrics, Emory University School of Medicine and Children's Healthcare of Atlanta, Atlanta, GA, E-mail: lisa.cranmer@ emory.edu. Judd L. Walson, Departments of Medicine, Pediatrics, Epidemiology, and Global Health, University of Washington, Seattle, WA, E-mail: walson@uw.edu.

REFERENCES

1. Murray CJ, Ortblad KF, Guinovart C, Lim SS, Wolock TM, Roberts DA, Dansereau EA, Graetz N, Barber RM, Brown JC, Wang H, Duber HC, Naghavi M, Dicker D, Dandona L, Salomon JA, Heuton KR, Foreman K, Phillips DE, Fleming TD, Flaxman AD, Phillips BK, Johnson EK, Coggeshall MS, Abd-Allah F, Abera SF, Abraham JP, Abubakar I, Abu-Raddad LJ, Abu-Rmeileh NM, Achoki T, Adeyemo AO, Adou AK, Adsuar JC, Agardh EE, Akena D, Al Kahbouri MJ, Alasfoor D, Albittar MI, Alcala-Cerra G, Alegretti MA, Alemu ZA, Alfonso-Cristancho R, Alhabib S, Ali R, Alla F, Allen PJ, Alsharif U, Alvarez E, Alvis-Guzman N, Amankwaa AA, Amare AT, Amini H, Ammar W, Anderson BO, Antonio CA, Anwari P, Arnlov J, Arsenijevic VS, Artaman A, Asghar RJ, Assadi R, Atkins LS, Badawi A, Balakrishnan K, Banerjee A, Basu S, Beardsley J, Bekele T, Bell ML, Bernabe E, Beyene TJ, Bhala N, Bhalla A, Bhutta ZA, Abdulhak AB, Binagwaho A, Blore JD, Basara BB, Bose D, Brainin M, Breitborde N, Castaneda-Orjuela CA, Catala-Lopez F, Chadha VK, Chang JC, Chiang PP, Chuang TW, Colomar M, Cooper LT, Cooper C, Courville KJ, Cowie BC, Criqui MH, Dandona R, Dayama A, De Leo D, Degenhardt L, Del Pozo-Cruz B, Deribe K, Des Jarlais DC, Dessalegn M, Dharmaratne SD, Dilmen U, Ding EL, Driscoll TR, Durrani AM, Ellenbogen RG, Ermakov SP, Esteghamati A, Faraon EJ, Farzadfar F, Fereshtehnejad SM, Fijabi DO, Forouzanfar MH, Fra Paleo U, Gaffikin L, Gamkrelidze A, Gankpé FG, Geleijnse JM, Gessner BD, Gibney KB, Ginawi IA, Glaser EL, Gona P, Goto A, Gouda HN, Gugnani HC, Gupta R, Gupta R, Hafezi-Nejad N, Hamadeh RR, Hammami M, Hankey GJ, Harb HL, Haro JM, Havmoeller R, Hay SI, Hedavati MT, Pi IB, Hoek HW, Hornberger JC, Hosgood HD, Hotez PJ, Hoy DG, Huang JJ, Iburg KM, Idrisov BT, Innos K, Jacobsen KH, Jeemon P, Jensen PN, Jha V, Jiang G, Jonas JB, Juel K, Kan H, Kankindi I, Karam NE, Karch A, Karema CK, Kaul A, Kawakami N, Kazi DS, Kemp AH, Kengne AP, Keren A. Kereselidze M. Khader YS. Khalifa SE. Khan EA. Khang YH, Khonelidze I, Kinfu Y, Kinge JM, Knibbs L, Kokubo Y, Kosen S, Defo BK, Kulkarni VS, Kulkarni C, Kumar K, Kumar RB, Kumar GA, Kwan GF, Lai T, Balaji AL, Lam H, Lan Q, Lansingh VC, Larson HJ, Larsson A, Lee JT, Leigh J, Leinsalu M, Leung R, Li Y, Li Y, De Lima GM, Lin HH, Lipshultz SE, Liu S, Liu Y, Lloyd BK, Lotufo PA, Machado VM, Maclachlan JH, Magis-Rodriguez C, Majdan M, Mapoma CC, Marcenes W, Marzan MB, Masci JR, Mashal MT, Mason-Jones AJ, Mayosi BM, Mazorodze TT, Mckay AC, Meaney PA, Mehndiratta MM, Mejia-Rodriguez F, Melaku YA, Memish ZA, Mendoza W, Miller TR, Mills EJ, Mohammad KA, Mokdad AH, Mola GL, Monasta L, Montico M, Moore AR, Mori R, Moturi WN, Mukaigawara M, Murthy KS, Naheed A, Naidoo KS, Naldi L, Nangia V, Narayan KM, Nash D, Nejjari C, Nelson RG, Neupane SP, Newton CR, Ng M, Nisar MI, Nolte S, Norheim OF, Nowaseb V, Nyakarahuka L, Oh IH, Ohkubo T, Olusanya BO, Omer SB, Opio JN, Orisakwe OE, Pandian JD, Papachristou C, Caicedo AJ, Patten SB, Paul VK, Pavlin BI, Pearce N, Pereira DM, Pervaiz A, Pesudovs K, Petzold M, Pourmalek F, Qato D, Quezada AD, Quistberg DA, Rafay A, Rahimi K, Rahimi-Movaghar V, Ur

Rahman S, Raju M, Rana SM, Razavi H, Reilly RQ, Remuzzi G, Richardus JH, Ronfani L, Roy N, Sabin N, Saeedi MY, Sahraian MA, Samonte GM, Sawhney M, Schneider IJ, Schwebel DC, Seedat S, Sepanlou SG, Servan-Mori EE, Sheikhbahaei S, Shibuya K, Shin HH, Shiue I, Shivakoti R, Sigfusdottir ID, Silberberg DH, Silva AP, Simard EP, Singh JA, Skirbekk V, Sliwa K, Soneji S, Soshnikov SS, Sreeramareddy CT, Stathopoulou VK, Stroumpoulis K, Swaminathan S, Sykes BL, Tabb KM, Talongwa RT, Tenkorang EY, Terkawi AS, Thomson AJ, Thorne-Lyman AL, Towbin JA, Traebert J, Tran BX, Dimbuene ZT, Tsilimbaris M, Uchendu US, Ukwaja KN, Uzun SB, Vallely AJ, Vasankari TJ, Venketasubramanian N, Violante FS, Vlassov VV, Vollset SE, Waller S, Wallin MT, Wang L, Wang X, Wang Y, Weichenthal S, Weiderpass E, Weintraub RG, Westerman R, White RA, Wilkinson JD, Williams TN, Woldeyohannes SM, Wong JQ, Xu G, Yang YC, Yano Y, Yentur GK, Yip P, Yonemoto N, Yoon SJ, Younis M, Yu C, Jin KY, El Sayed Zaki M, Zhao Y, Zheng Y, Zhou M, Zhu J, Zou XN, Lopez AD, Vos T, 2014. Global, regional, and national incidence and mortality for HIV, tuberculosis, and malaria during 1990-2013: a systematic analysis for the Global Burden of Disease Study 2013. Lancet 384: 1005-1070.

- Jenkins HE, Tolman AW, Yuen CM, Parr JB, Keshavjee S, Perez-Velez CM, Pagano M, Becerra MC, Cohen T, 2014. Incidence of multidrug-resistant tuberculosis disease in children: systematic review and global estimates. *Lancet* 383: 1572–1579.
- Stop TB Partnership Childhood TB Subgroup World Health Organizaiton, 2006. Guidance for National Tuberculosis Programmes on the management of tuberculosis in children. Chapter 1: introduction and diagnosis of tuberculosis in children. *Int J Tuberc Lung Dis 10*: 1091–1097.
- Marais BJ, 2011. Childhood tuberculosis: epidemiology and natural history of disease. *Indian J Pediatr* 78: 321–327.
- Archibald LK, den Dulk MO, Pallangyo KJ, Reller LB, 1998. Fatal *Mycobacterium tuberculosis* bloodstream infections in febrile hospitalized adults in Dar es Salaam, Tanzania. *Clin Infect Dis 26:* 290–296.
- Gilks CF, Brindle RJ, Mwachari C, Batchelor B, Bwayo J, Kimari J, Arbeit RD, von Reyn CF, 1995. Disseminated *Mycobacterium avium* infection among HIV-infected patients in Kenya. J Acquir Immune Defic Syndr Hum Retrovirol 8: 195–198.
- Arthur G, Nduba VN, Kariuki SM, Kimari J, Bhatt SM, Gilks CF, 2001. Trends in bloodstream infections among human immunodeficiency virus-infected adults admitted to a hospital in Nairobi, Kenya, during the last decade. *Clin Infect Dis 33:* 248–256.
- Bell M, Archibald LK, Nwanyanwu O, Dobbie H, Tokars J, Kazembe PN, Reller LB, Jarvis WR, 2001. Seasonal variation in the etiology of bloodstream infections in a febrile inpatient population in a developing country. *Int J Infect Dis* 5: 63–69.
- Crump JA, Ramadhani HO, Morrissey AB, Saganda W, Mwako MS, Yang LY, Chow SC, Njau BN, Mushi GS, Maro VP, Reller LB, Bartlett JA, 2012. Bacteremic disseminated tuberculosis in sub-saharan Africa: a prospective cohort study. *Clin Infect Dis* 55: 242–250.
- Jacob ST, Pavlinac PB, Nakiyingi L, Banura P, Baeten JM, Morgan K, Magaret A, Manabe Y, Reynolds SJ, Liles WC, Wald A, Joloba ML, Mayanja-Kizza H, Scheld WM, 2013. *Mycobacterium tuberculosis* bacteremia in a cohort of HIV-infected patients hospitalized with severe sepsis in Uganda-high frequency, low clinical suspicion [corrected] and derivation of a clinical prediction score. *PLoS One 8:* e70305.
- 11. Ssali FN, Kamya MR, Wabwire-Mangen F, Kasasa S, Joloba M, Williams D, Mugerwa RD, Ellner JJ, Johnson JL, 1998. A prospective study of community-acquired bloodstream infections among febrile adults admitted to Mulago Hospital in Kampala, Uganda. J Acquir Immune Defic Syndr Hum Retrovirol 19: 484–489.
- 12. Crump JA, Wu X, Kendall MA, Ive PD, Kumwenda JJ, Grinsztejn B, Jentsch U, Swindells S, 2015. Predictors and outcomes of *Mycobacterium tuberculosis* bacteremia among patients with HIV and tuberculosis co-infection enrolled in the ACTG A5221 STRIDE study. *BMC Infect Dis* 15: 12.
- Gray KD, Cunningham CK, Clifton DC, Afwamba IA, Mushi GS, Msuya LJ, Crump JA, Buchanan AM, 2013. Prevalence

of mycobacteremia among HIV-infected infants and children in northern Tanzania. *Pediatr Infect Dis J* 32: 754–756.

- Archibald LK, Kazembe PN, Nwanyanwu O, Mwansambo C, Reller LB, Jarvis WR, 2003. Epidemiology of bloodstream infections in a bacille Calmette-Guerin-vaccinated pediatric population in Malawi. J Infect Dis 188: 202–208.
- Waddell RD, Lishimpi K, von Reyn CF, Chintu C, Baboo KS, Kreiswirth B, Talbot EA, Karagas MR, Dartmouth UUCSG, 2001. Bacteremia due to *Mycobacterium tuberculosis* or M. bovis, Bacille Calmette-Guerin (BCG) among HIV-positive children and adults in Zambia. *AIDS* 15: 55–60.
- National AIDS, Control Programme STI (NASCOP), 2014. Kenya AIDS Indicator Survey 2012: Final Report. Nairobi, Kenya: NASCOP.
- 17. van't Hoog AH, Laserson KF, Githui WA, Meme HK, Agaya JA, Odeny LO, Muchiri BG, Marston BJ, DeCock KM, Borgdorff MW, 2011. High prevalence of pulmonary tuberculosis and inadequate case finding in rural western Kenya. *Am J Respir Crit Care Med 183*: 1245–1253.
- Lee LH, LeVea CM, Graman PS, 1998. Congenital tuberculosis in a neonatal intensive care unit: case report, epidemiological investigation, and management of exposures. *Clin Infect Dis* 27: 474–477.
- WHO Multicentre Growth Reference Study Group, 2006. WHO Child Growth Standards: Length/Height-for-Age, Weight-for-Age, Weight-for-Length, Weight-for-Height and Body Mass Indexfor-Age: Methods and Development. Geneva, Switzerland: World Health Organization, 312. Available at: http://www.who.int/child growth/publications/en/).
- Onchiri FM, Pavlinac PB, Singa BO, Naulikha JM, Odundo EA, Farquhar C, Richardson BA, John-Stewart G, Walson JL, 2015. Low bacteremia prevalence among febrile children in areas of differing malaria transmission in rural Kenya: a crosssectional study. *J Pediatric Infect Dis Soc* (Published online ahead of print). doi:10.1093/jpids/piv043.
- Kenya National Bureau of Statistics, 2013. Nyanza Province Multiple Indicator Cluster Survey 2011, Final Report. Nairobi, Kenya: Kenya National Bureau of Statistics.
- 22. Center for Disease Control and Prevention, 2015. *Tuberculosis in Specific Populations-Children*. Available at: http://www.cdc .gov/tb/topic/populations/tbinchildren/default.htm.
- Isaacman DJ, Karasic RB, Reynolds EA, Kost SI, 1996. Effect of number of blood cultures and volume of blood on detection of bacteremia in children. J Pediatr 128: 190–195.
- Cockerill FR 3rd, Wilson JW, Vetter EA, Goodman KM, Torgerson CA, Harmsen WS, Schleck CD, Ilstrup DM, Washington JA 2nd, Wilson WR, 2004. Optimal testing parameters for blood cultures. *Clin Infect Dis* 38: 1724–1730.
- 25. Heysell SK, Thomas TA, Gandhi NR, Moll AP, Eksteen FJ, Coovadia Y, Roux L, Babaria P, Lalloo U, Friedland G, Shah S, 2010. Blood cultures for the diagnosis of multidrug-resistant and extensively drug-resistant tuberculosis among HIV-infected patients from rural South Africa: a cross-sectional study. BMC Infect Dis 10: 344.

- Bacha HA, Cimerman S, de Souza SA, Hadad DJ, Mendes CM, 2004. Prevalence of mycobacteremia in patients with AIDS and persistant fever. *Braz J Infect Dis* 8: 290–295.
- 27. Graham SM, Ahmed T, Amanullah F, Browning R, Cardenas V, Casenghi M, Cuevas LE, Gale M, Gie RP, Grzemska M, Handelsman E, Hatherill M, Hesseling AC, Jean-Philippe P, Kampmann B, Kabra SK, Lienhardt C, Lighter-Fisher J, Madhi S, Makhene M, Marais BJ, McNeeley DF, Menzies H, Mitchell C, Modi S, Mofenson L, Musoke P, Nachman S, Powell C, Rigaud M, Rouzier V, Starke JR, Swaminathan S, Wingfield C, 2012. Evaluation of tuberculosis diagnostics in children: 1. Proposed clinical case definitions for classification of intrathoracic tuberculosis disease. Consensus from an expert panel. J Infect Dis 205 (Suppl 2): S199–S208.
- Dubey A, Gwal R, Agrawal S, 2013. Mycobacterium tuberculosis detection in blood using multiplex nested polymerase chain reaction. Int J Tuberc Lung Dis 17: 1341–1345.
- Rebollo MJ, San Juan Garrido R, Folgueira D, Palenque E, Diaz-Pedroche C, Lumbreras C, Aguado JM, 2006. Blood and urine samples as useful sources for the direct detection of tuberculosis by polymerase chain reaction. *Diagn Microbiol Infect Dis* 56: 141–146.
- 30. Feasey NA, Banada PP, Howson W, Sloan DJ, Mdolo A, Boehme C, Chipungu GA, Allain TJ, Heyderman RS, Corbett EL, Alland D, 2013. Evaluation of Xpert MTB/RIF for detection of tuberculosis from blood samples of HIVinfected adults confirms *Mycobacterium tuberculosis* bacteremia as an indicator of poor prognosis. *J Clin Microbiol 51:* 2311–2316.
- Colditz GA, Berkey CS, Mosteller F, Brewer TF, Wilson ME, Burdick E, Fineberg HV, 1995. The efficacy of bacillus Calmette-Guerin vaccination of newborns and infants in the prevention of tuberculosis: meta-analyses of the published literature. *Pediatrics 96*: 29–35.
- Rodrigues LC, Diwan VK, Wheeler JG, 1993. Protective effect of BCG against tuberculous meningitis and miliary tuberculosis: a meta-analysis. *Int J Epidemiol 22*: 1154–1158.
- Cohn DL, 1997. Use of the bacille Calmette-Guerin vaccination for the prevention of tuberculosis: renewed interest in an old vaccine. *Am J Med Sci 313*: 372–376.
- Trunz BB, Fine P, Dye C, 2006. Effect of BCG vaccination on childhood tuberculous meningitis and miliary tuberculosis worldwide: a meta-analysis and assessment of cost-effectiveness. *Lancet* 367: 1173–1180.
- 35. Marais BJ, Gie RP, Hesseling AC, Schaaf HS, Lombard C, Enarson DA, Beyers N, 2006. A refined symptom-based approach to diagnose pulmonary tuberculosis in children. *Pediatrics 118:* e1350–e1359.
- Mwinga A, 2005. Challenges and hope for the diagnosis of tuberculosis in infants and young children. *Lancet* 365: 97–98.
- Dodd PJ, Gardiner E, Coghlan R, Seddon JA, 2014. Burden of childhood tuberculosis in 22 high-burden countries: a mathematical modelling study. *Lancet Glob Health 2:* e453–e459.